

1 **Abundance and genetic damage of barn swallows from**
2 **Fukushima**

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26 A. BONISOLI-ALQUATI ET AL. BARN SWALLOWS AFTER FUKUSHIMA

27 **Abstract**

28 A number of studies have assessed or modeled the distribution of the radionuclides
29 released by the accident at the Fukushima-Daiichi Nuclear Power Plant (FDNPP). Few studies
30 however have investigated its consequences for the local biota.

31 We tested whether exposure of barn swallow (*Hirundo rustica*) nestlings to low dose
32 ionizing radiation increased genetic damage to their peripheral erythrocytes. We estimated
33 external radiation exposure by using thermoluminescent dosimeters, and by measuring
34 radioactivity of the nest material. We then assessed DNA damage by means of the neutral comet
35 assay. In addition, we conducted standard point-count censuses of barn swallows across
36 environmental radiation levels, and estimated their abundance and local age ratio.

37 Radioactivity of nest samples was in the range 479–143,349 Bq kg⁻¹, while external
38 exposure varied between 0.15 and 4.9 mGy. Exposure to radioactive contamination did not
39 correlate with higher genetic damage in nestlings. However, at higher levels of radioactive
40 contamination the number of barn swallows declined and the fraction of juveniles decreased,
41 indicating lower survival and lower reproduction and/or fledging rate.

42 Thus, genetic damage to nestlings does not explain the decline of barn swallows in
43 contaminated areas, and a proximate mechanism for the demographic effects documented here
44 remains to be clarified.

45
46 *Keywords:* DNA damage; Fukushima-Daiichi; ionizing radiation; nuclear disaster; radioactive
47 contamination; Tohoku Earthquake

48

49 Introduction

50 On March 11 2011, a tsunami caused by the Great East Japan Earthquake seriously
51 damaged the electrical and the cooling systems of the Fukushima-Daiichi Nuclear Power Plant
52 (FDNPP), causing hydrogen explosions at the Unit 1, 2 and 3 reactors. These explosions released
53 large amounts of high volatility fission products, including ^{129}mTe , ^{131}I , ^{133}Xe , ^{134}Cs , ^{136}Cs , and
54 ^{137}Cs ¹⁻³. Although the estimates of the release vary considerably⁴⁻¹⁰, the accident is universally
55 regarded as the second largest release of radionuclides in history after the Chernobyl accident,
56 with estimates of total radioactivity released in the range of up to hundreds of PBq¹⁰. Such
57 massive discharge of radionuclides raises concern about its possible consequences for
58 environmental and human health¹¹, particularly given the persistence of ^{137}Cs in the
59 environment.

60 Predictably, large efforts have since been devoted to model the atmospheric release, the
61 deposition of radionuclides and their redistribution^{2,12}. Several other studies have assessed the
62 concentration of the radionuclides in the biological tissues of animals and plants (mammals:^{13,14};
63 fish:^{7,15}; birds:¹⁶; plants¹⁷⁻¹⁹).

64 Few studies so far have examined the potential biological consequences of exposure to
65 radionuclides released by the accident. A study on the pale blue grass butterfly (*Zizeeria maha*)
66 that coupled field sampling and rearing of individuals under common garden conditions showed
67 an increase in aberrations in the coloration and patterns of wings^{20,21}. A study on earthworms
68 also demonstrated that animals from sites where radiation level was as low as 2.8 $\mu\text{Sv/h}$ had
69 higher DNA damage than animals from control sites²². A recent study of wild Japanese
70 macaques (*Macaca fuscata*) found that individuals from Fukushima had lower white blood cell
71 (WBCs) and red blood cell counts (RBCs), lower hemoglobin concentration and lower
72 hematocrit values than those sampled in the Shimokita peninsula, at a distance of 400 km from
73 the FDNPP²³. Vitamin A levels of streaked shearwaters (*Calonectris leucomelas*) sampled in
74 colonies exposed to contamination from the FDNPP were lower than in animals from a colony
75 that was not reached by the plume²⁴.

76 Ecological studies conducted in the Chernobyl Exclusion Zone have indicated that
77 radiation levels comparable to those found around Fukushima can be associated with deleterious
78 genetic, physiological and life-history consequences for exposed wildlife²⁵. Low-dose radiation
79 in the Chernobyl region was associated with higher DNA damage in adult barn swallows²⁶,

80 higher frequency of morphological abnormalities and tumors ^{27,28}, and a reduction in brain size ²⁹.
81 These and other physiological and genetic consequences of radiation exposure in Chernobyl ³⁰
82 have been indicated as the likely cause underlying the higher mortality and the populations
83 declines of many bird species living in the Chernobyl region, as inferred from point count
84 censuses ³¹, and age ratios from mist netting studies ³².

85 In spite of differences between the two accidents in the quality and amount of
86 contaminants scattered and the number of generations of exposure, early studies suggest that
87 similarities also exist in the response of natural populations to radioactive contamination. Point-
88 count censuses conducted around Fukushima in 2011 have found that bird population in
89 radioactively contaminated areas have declined similarly as in Chernobyl ³³. Later surveys
90 validated this finding and concluded that the contamination might have had an even larger
91 negative effect during 2012 ³⁴.

92 Here, we describe the results of a study on barn swallow nestlings during May-June 2012
93 to investigate whether exposure to radiation is affecting their genetic integrity prior to fledging.
94 We estimated external radiation exposure of nestlings by attaching thermoluminescent
95 dosimeters (TLDs) to their nest, and by collecting a sample of nest material whose activity
96 concentration we measured using gamma spectrometry in the lab.

97 We also describe the results of a survey of barn swallows that we conducted in July 2011-
98 2013 across gradients of radioactive contamination spanning almost two orders of magnitude.
99 Part of this database (2011 and 2012) has been previously published in studies relating the
100 abundance of birds in the Fukushima region to the level of radioactive contamination ^{33,34}. In
101 addition to presenting an additional year of data, here we focus the analyses on the local
102 abundance of barn swallows. We also present an analysis of the age ratio of barn swallows,
103 which can be readily determined from plumage characteristics, predicting that higher levels of
104 radiation would lead to a lower fraction of juveniles due to egg infertility and death of nestlings
105 ³⁵.

106 With a few notable exceptions ^{16,23,24}, all studies conducted so far have at most analyzed the
107 concentrations of radioisotopes in the tissues of organisms, but neglected the assessment of
108 markers of their potential biological effects. The results that we describe represent the first
109 extensive investigation of the potential genotoxicity of measured radiation exposure in any wild
110 population of birds from the Fukushima region.

111

112 **Results**113 *Radioactivity of nest samples and radiation exposure of nestlings*

114 The average exposure measured by the TLDs was 0.59 mGy (0.79 mGy SD; range: 0.15 –
 115 4.9 mGy; N = 43), corresponding to an average dose rate of 0.90 $\mu\text{Gy h}^{-1}$ (1.24 $\mu\text{Gy h}^{-1}$ SD;
 116 range: 0.23 – 7.52 $\mu\text{Gy h}^{-1}$).

117 The activity concentrations measured in the nest samples were 10,730 Bq kg^{-1} dry weight
 118 (d.w.) (18,276 SD; range 318–82,409 Bq kg^{-1} d.w.; N = 45) for ^{137}Cs , and 8,656 Bq kg^{-1} d.w.
 119 (14,433 SD; range 128–60,940 Bq kg^{-1} d.w.; N = 45) for ^{134}Cs . When we combined the activities
 120 measured for each radionuclide in a single estimate, the total radioactivity was 19,386 Bq kg^{-1}
 121 d.w. (32,681 SD; range 478–143,349 Bq kg^{-1} d.w.; N = 45). Total radioactivity of the nest
 122 material significantly positively predicted the radiation dose received by the TLDs ($t_{39} = 6.74$, p
 123 < 0.0001 , $R^2 = 0.54$, N = 40; Supplementary Fig. 1). Environmental radiation levels significantly
 124 positively correlated with the dose received by the TLDs ($t_{42} = 4.88$, $p < 0.0001$, $R^2 = 0.37$, N=
 125 43; Supplementary Fig. 1). Environmental radiation levels also positively correlated with the
 126 specific activity of ^{137}Cs ($t_{44} = 2.22$, $P = 0.032$, $R^2 = 0.08$, N= 45), and of ^{134}Cs ($t_{44} = 2.37$, $P =$
 127 0.022 , $R^2 = 0.09$, N= 45).

128

129 *Radiation exposure and genetic damage of nestlings*

130 The average DNA damage, as indexed by the percentage of DNA in the comet tail, was
 131 10.04 (4.86 SD; range: 2.83-23.41). The total activity concentration of the nest material, obtained
 132 by combining the estimates for ^{134}Cs and ^{137}Cs , did not significantly predict genetic damage of
 133 nestlings ($F_{1,624} = 0.51$, $p = 0.502$, N = 49, Fig. 1a), nor did the dose to the TLDs attached to the
 134 nest ($F_{1,9,81} = 0.33$, $p = 0.577$, N = 49, Fig. 1b). We tested if there was a difference in variance in
 135 DNA damage at high radiation levels by splitting our dataset in two groups using median
 136 radioactivity of the nest sample or median dose to the TLDs as the cutting points. Variance in
 137 DNA damage did not differ significantly between nestlings from more radioactive nests and
 138 those from less radioactive nests (Levene's test: $F_{1,47} = 0.015$, $p = 0.904$). Similarly, variance in
 139 DNA damage did not differ significantly between nests where the TLDs received a higher dose
 140 and those where the dose to the TLDs was lower (Levene's test: $F_{1,47} = 0.039$, $p = 0.844$).

141 In none of these analyses did body mass and estimated age of the nestling significantly
142 predict genetic damage of the nestlings (see Table 1 in Electronic Supplementary Material). The
143 effect of the nest of origin was never significant in predicting genetic damage of the nestlings
144 (Table 1 in Electronic Supplementary Materials).

145

146 *Abundance and age ratio of barn swallows*

147 Radiation levels at the breeding bird census points ranged from 0.18 to 38.11 $\mu\text{Sv/h}$ [mean
148 (SD) = 7.16 $\mu\text{Sv/h}$ (7.90), $N = 1100$]. The abundance of barn swallows, as inferred from our
149 point count censuses, significantly declined with increasing environmental radiation levels
150 ($F_{1,1093} = 105.81, p < 0.0001$; slope (SE) = -1.18 (0.12); Fig. 2]. The number of barn swallows
151 increased with increasing farmland ($F_{1,1093} = 12.53, p = 0.0004$; slope (SE) = 6.82×10^{-3} ($1.37 \times$
152 10^{-3})), and decreased with increasing ground coverage by grass ($F_{1,1093} = 320.78, p < 0.0001$;
153 slope (SE) = -4.75×10^{-2} (2.18×10^{-3})) and coniferous forest ($F_{1,1093} = 4.98, p = 0.0256$; slope
154 (SE) = -1.74 (0.39)). In addition, the abundance of barn swallows differed among years ($F_{2,1093} =$
155 $103.83, p < 0.0001$). There were significantly fewer barn swallows in 2012 than in 2011 ($t =$
156 $23.1, p < 0.0001$) and fewer in 2013 compared to 2012 ($t = 7.85, p = 0.005$), or the two previous
157 years combined ($t = 34.07, p < 0.0001$).

158 The probability of a barn swallow being a juvenile decreased significantly with increasing
159 environmental radiation levels ($F_{1,1069} = 13.50, p = 0.0002$; slope (SE) = -1.84 (0.57); Fig. 3]. In
160 addition, juvenile barn swallows were more common where adults were more abundant, as
161 expected from the fact that the adults produced the offspring ($F_{1,1069} = 32.84, p < 0.0001$; slope
162 (SE) = 1.00 (0.18). There was also a significant variation among years in the probability that a
163 bird was a juvenile ($F_{2,1069} = 15.44, p = 0.021$). This probability was higher in 2012 than in 2011
164 ($t = 6.04, p = 0.014$), or 2013 ($t = 6.04, p = 0.014$).

165

166 **Discussion**

167 In this study, we investigated genetic damage in barn swallows nestlings exposed to
168 radioactive contamination following the accident at the Fukushima Daiichi Nuclear Power Plant
169 in March 2011. We also estimated the abundance of barn swallows across sites differing in
170 environmental radiation levels by almost two orders of magnitude, while also assessing the
171 relative frequency of juveniles and adults. To the best of our knowledge, this is the first study at

172 Fukushima relating a known biomarker of radiation exposure to estimates of radiation exposure
173 in any wild population of animals exposed to the radioactive fallout.

174 We could not detect any increase in genetic damage in nestlings exposed to a range of
175 contamination levels during their rearing period. These results partially conflict with previous
176 results in adult barn swallows from the Chernobyl region, where higher genetic damage was
177 demonstrated at levels that were comparable to the ones detected in this study²⁶. It could be
178 argued that barn swallow nestlings at Fukushima were exposed for shorter times compared to
179 adults in Chernobyl. The exposure period of barn swallow nestlings (averaging 26 ± 5 days in
180 our sample, due to the combined duration of the incubation period and the rearing period) is
181 considerably shorter compared to the months-long residence of adult barn swallows at the
182 breeding sites. Alternatively, differences in the mixture of radionuclides scattered by the two
183 disasters could explain the difference in the effect, if different mixtures have different associated
184 risks, due to differences in particle emission. While ^{134}Cs and ^{137}Cs are the dominant
185 radionuclides dispersed by the Fukushima disaster¹², ^{137}Cs , ^{90}Sr , ^{241}Am and several radioisotopes
186 of plutonium are the ones currently present around the Chernobyl Exclusion Zone³⁶. The greater
187 abundance of Pu isotopes and other actinides in Chernobyl could thus be responsible for the
188 difference between the two disasters, as alpha emitters have large health effects. Finally,
189 differences in historical exposure and associated trans-generational accumulations of deleterious
190 effects could be responsible for the differences between the two disasters. Swallow populations
191 in the Chernobyl region have been chronically exposed to radioactive contamination for over 20
192 years at the time of sampling for our 2010 study of genetic damage. Conversely, exposure to
193 radioactive contamination only lasted little more than one year when we sampled barn swallows
194 in the contaminated areas around Fukushima for the present study. While this difference in
195 historical exposure is expected to affect mutation accumulation, predictions regarding the
196 resistance of natural populations to radiation-induced genetic damage are less clear, as genetic
197 damage is not inherited, and natural populations are expected to evolve resistance to radiation-
198 induced oxidative damage over generations³⁷. Given that the nestlings examined here belonged
199 to the second generation after the disaster, and the first one from parents that were themselves
200 exposed throughout development, we do not expect the lack of an increase in genetic damage to
201 be due to selection for adaptation to ionizing radiation.

202 At present, the interpretation of the variation among nestlings in their levels of DNA
203 damage is deemed to be largely speculative. Differences in individual growth rates or in the
204 intensity of competition with siblings could account for such variation through an effect on the
205 oxidative status of nestlings. Consistent with this, resistance of red blood cells to free radicals
206 has been found to negatively correlate with growth rate in zebra finch (*Taeniopygia guttata*)
207 nestlings³⁸. In addition, magpie (*Pica pica*) nestlings begging more intensively were shown to
208 have higher levels of lipid peroxidation, as indexed by their malondialdehyde levels³⁹. Future
209 studies where nestlings are repeatedly measured will allow controlling for these confounding
210 factors, uncovering potential more subtle effects of radiation.

211 Due to restriction in access to more highly contaminated areas during our sampling of
212 nestlings, we could not access sites where nestlings might have received considerably higher
213 doses, including the towns of Okuma, Futaba and Namie. Thus, the data presented here should
214 be cautiously interpreted when addressing whether exposure to radioactive contamination is
215 causing an increase in genetic damage in wild populations of animals in contaminated areas, as
216 higher contamination levels might imply more deleterious consequences. The barn swallows is a
217 model species for investigating the effects of radioactive contamination in Fukushima, due to its
218 abundance, philopatry to a once chosen breeding site, and availability of control as well as
219 affected populations (e.g. in Chernobyl). However, different species may vary in their
220 radiosensitivity and the lack of an effect in one species does not necessarily imply that all others
221 are similarly unaffected^{1-3,30,31,33,34}.

222 In addition, while the biomarker that we assessed did not show any response among
223 nestlings, our census of barn swallows confirmed previous findings of population declines of
224 several bird species in the Fukushima region^{33,34}. In addition, it suggested that the population
225 decline is due to lower fecundity and/or lower fledging rate, as demonstrated by a decrease in the
226 proportion of juveniles at higher levels of radiation exposure. This result is consistent with the
227 demonstrated decline in fertility, reproductive function and parental care that we have shown in
228 Chernobyl, in the barn swallow as well as in other species^{35,40-42}.

229 Human absence from highly contaminated towns, with the associated changes in the
230 farming practice and the lack of deterrence for the natural predators of this species (e.g. the
231 Japanese jungle crow *Corvus macrorhynchos*) is a potential alternative explanation for the
232 decline of this species in contaminated areas. In future studies, the assessment of biomarkers of

233 radiation exposure will help determine whether the decline of this species is due to a direct or
234 indirect effect of ionizing radiation (i.e. through an effect on human presence).

235 The discrepancy between the decline in abundance of barn swallows and the lack of any
236 response in the biomarker of genetic damage that we assessed in barn swallow nestlings call for
237 further investigation of the potential mechanistic (i.e. physiological and genetic) links between
238 radiation exposure and population dynamics. Multiple cytogenetic biomarkers of radiation
239 exposure will have to be investigated in the future, while at the same time expanding research
240 into areas contaminated to a higher degree than it was assessed in the present study. Similarly,
241 aural and visual censuses of diversity and abundance will have to be complemented with mist
242 netting of birds in order to estimate transfer of radionuclides to birds. It should be noted that
243 radiation levels examined during the censuses were much wider than the range of contamination
244 levels where nestlings have been measured, as more highly contaminated areas could not be
245 sampled due to lack of access or sampling permit. Thus, the present results should not be
246 interpreted as indicating that no deleterious consequence is expected over the entire area that was
247 contaminated by the radioactive fallout, nor should they be taken as evidence that genetic
248 damage at the *adult* stage is not mediating the population decline of barn swallows, as adult birds
249 were not assessed in the present study. In fact, the investigation of genetic damage of nestlings
250 should also be expanded to more highly contaminated areas in order to exclude the possibility
251 that it contributed to the decline of barn swallow populations.

252 Overall, our radioactivity measurements are compatible with previously published
253 measurements and dose estimates^{43,44}. Higher exposure levels for barn swallows can thus be
254 predicted in more highly contaminated areas. The exposure levels measured here are consistent
255 with the occurrence of physiological and life-history consequences (i.e. reduced survival and
256 reproduction) in exposed organisms⁴⁴. A recent analysis that inferred doses from published
257 information on contamination levels and used official benchmarks for dose-response also
258 concluded that exposure to contamination following the accident could induce sub-lethal effects
259 on the populations of terrestrial vertebrates⁴⁵. This same analysis, however, also concluded that
260 any population-level consequence of such individual-level doses would be unlikely, thus raising
261 the issue of reconciling measured doses with population declines that have been shown by recent
262 censuses^{33,34}. At the same time, it should be noted that our estimates of radiation exposure are
263 conservative, as they admittedly do not account for internal radiation exposure due to inhalation

264 or ingestion of radionuclides. Dose conversion coefficients (DCC) for internal exposure to ^{134}Cs
265 and ^{137}Cs in species ecologically similar to the barn swallow are expected to be at least as large
266 as the DCCs for external exposure ⁴⁶. Thus, future studies will have to improve dosimetry by
267 assessing both internal and external radiation exposure of local populations of barn swallows
268 potentially impacted by the fallout.

269 Similar population declines at levels of contamination that are not predicted to have
270 population-level consequences have also been observed in the Chernobyl region ^{31,32,47},
271 prompting similar skepticism ⁴⁸. There is increasing evidence, however, that the benchmarks
272 indicated as safe by international organizations (IAEA, ICRP)^{49,50} might be underestimating the
273 risk associated with exposure to ionizing radiation in the natural environment, especially under
274 chronic exposure ⁵¹. Hazards to natural populations have recently been found to arise at doses
275 considerably lower than it had been shown in controlled experiments in the lab ⁵¹. In addition, a
276 recent meta-analysis that reviewed studies conducted in very high background radiation areas
277 where radionuclides occur naturally found a consistent positive relationship between
278 environmental radiation levels and mutation rate, DNA repair and genetics, in human as well as
279 animal populations ⁵². The likely explanation for the discrepancy between the lab and an
280 ecologically-meaningful setting is that lab conditions are far more benign than realistic
281 ecological conditions, where food and essential nutrients are scarcer, predators and parasites are
282 more frequent, and other stressors may make the effects of ionizing radiation more apparent.

283

284 **Methods**

285 During May 2012, we attached thermoluminescent dosimeters (TLDs) to the inner and
286 outer rim of 55 barn swallow nests from the Fukushima region (Fig. 4). We used individually
287 calibrated LiF:Mg,Cu,P TLDs ($3.2 \times 3.2 \times 0.8$ mm; GR-200A), which have a higher sensitivity
288 than GR-100 TLDs ⁵³. The linearity and dose-response of the TLDs were measured with beams
289 produced by a medical linac and a ^{137}Cs source. We read the TLD response with a System 310
290 TLD Reader (Teledyne Brown Engineering), in a temperature range from room temperature to
291 240°C , at a rate of 10°C/s . The readings were consistent with previously published results ⁵⁴.
292 After an average 28.4 days (0.4 SE; range: 25-33 days), we retrieved the TLDs from the nest. On
293 the occasion of retrieving the TLDs, we collected a sample (~ 1 g) of nest material from the rim
294 of the nest. From 62 chicks from 16 nests that we estimated to be at least 7-8 days old, we also

295 collected a small blood sample (~50 μL), through puncture of the brachial vein and collection in
296 a heparinized capillary tube. We also transferred a drop of blood (~10 μL) to a vial containing
297 RNAprotect (Qiagen).

298 All procedures were performed in accordance with relevant guidelines and regulations, and
299 approved by the Institutional Animal Care and Use Committee (IACUC) of the University of
300 South Carolina (Protocol number: 2014-100237-052611).

301

302 *Radioactivity measurements*

303 In the field, we measured environmental α , β and γ radiation at the ground level below the
304 nest using a hand-held dosimeter (Model: Inspector, SE International, Inc., Summertown, TN,
305 USA).

306 We measured the activity concentrations of nest samples by conducting gamma ray
307 spectrometry with a SAM 940 Radioisotope Identifier (Berkeley Nucleonics, San Rafael, CA)
308 equipped with a 7.62×7.62 cm (3" \times 3") sodium iodide (NaI) detector. The spectrometer was
309 placed vertically within a lead detector shield (Canberra Industries, Meriden, CT, USA), with
310 additional shielding provided by lead bricks. We measured each sample by placing it on top of
311 the SAM 940 Radioisotope Identifier.

312 We later converted the spectra to activity measurements after calibration of the instrument
313 using standard ^{137}Cs and ^{134}Cs sources. For analysis we focused on the 661 keV decay gamma
314 from ^{137}Cs and the 597 and 796 keV peaks from ^{134}Cs . The samples were dried in a heating oven
315 with mechanical convection (Binder Inc., Bohemia, NY) at the temperature of 60°C for 12 h, and
316 weighed using a Sartorius electronic balance (Model R160P; Göttingen, Germany).

317 A high statistics, "empty target" spectrum was collected prior to the sample readings and
318 subtracted from all spectra to remove counts not associated with radioactive decay from the
319 sample. A linear background function was then fit to the peak region (490-500 keV) to remove
320 the continuum gammas and isolate the decay peaks. The 597 keV peak from ^{134}Cs and the 661
321 keV peak from ^{137}Cs overlapped considerably while the 796 keV peak from ^{134}Cs was resolved
322 completely. A spectrum from the ^{134}Cs calibration source was normalized to the data spectrum by
323 fitting to the 796keV peak. This fit was then subtracted from the entire sample spectrum to
324 isolate the 661 keV peak from ^{137}Cs . Integrating and comparing the counts in the decay peaks
325 from the samples to the counts in the same peaks from the known calibration sources produced

326 the absolute calibration. The total activity of each sample was calculated by summing the
327 activities estimated for ^{137}Cs and ^{134}Cs .

328 To estimate total duration of exposure for each nestling, we summed the estimated age of
329 the nestlings and the duration of the incubation period, which we conservatively estimated at 14
330 days.

331

332 *Analysis of genetic damage*

333 We estimated genetic damage using a single cell gel electrophoresis assay, also known as
334 comet assay, following the protocol reported in ⁵⁵, with minor modifications.

335 We prepared slides in advance by dipping single-frosted slides (VWR, Radnor, PA) in 1.5
336 % normal melting-point agarose. We transferred 3 μL of the solution of blood in RNAProtect
337 (Qiagen) to 997 μL of 1x PBS. We then mixed 50 μL of the solution with 450 μL of 1.5% low
338 melting-point agarose, and layered 100 μL of this mixture on the slides, covering with a glass
339 coverslip. We allowed the agarose to solidify for five minutes at 4°C. We then removed the
340 coverslip and added another layer of 100 μL of low melting-point agarose, and again allowed to
341 solidify for five minutes, before removing the coverslip. The slides were left for 1 hour at 4°C to
342 allow the solidification of the gel, and then immersed in cold lysis buffer (1% sodium
343 sarcosinate, 2.5 M NaCl, 100 mM Na₂EDTA, 10 mM Tris, 1% Triton X-100 added immediately
344 prior to use, at a final pH = 10), where they were kept for 1 hour at 4°C. We then rinsed the
345 slides with cold ddH₂O, and immersed them in neutral buffer (300 mM NaOH, 100 mM Tris, pH
346 10.0), to allow unwinding of the DNA for 30 minutes at 4°C. We electrophoresed slides in a tank
347 filled with the same buffer for 30 minutes at 0.7 V/cm and 150 mA at 4°C. After electrophoresis,
348 we rinsed the slides in a neutralization buffer (0.4M Tris, pH 7.4) three times, for five minutes
349 each. The slides were then fixed in 70% ethanol for 15 minutes, and left to dry overnight. We ran
350 four slides per each individual.

351 We stained the slides by immersion in a 1:10,000 solution of SYBR[®] Gold (Trevigen,
352 Gaithersburg, MD) for five minutes. Slides were then de-stained through immersion in a bath of
353 dd-H₂O for five minutes, and left to dry. Images of individual cells were captured using a
354 Metafer System (Metasystems, Bethesda, MD), an automated system that performs detection and
355 scoring of individual cells ⁵⁶. Only the nestlings for whom we had captured at least 100 cells
356 across the different slides were retained in the final sample ⁵⁷. In the final analyses we included

357 49 nestlings belonging to 16 nests, representing 78% of the initial sample of 63 nestlings. On
358 average we captured 313 cells (147 SD; range: 111-725) from 1-4 slides. As a measure of
359 damage to the DNA we used the percentage of DNA in the tail, which is a measure based on the
360 relative fluorescence intensity of the tail compared to the head of the comet, an the most reliable
361 parameter for the comet assay^{57,58}.

362 Data on genetic damage in each nestling were obtained by averaging percentages of DNA
363 in the tail of the comet across all cells.

364

365 *Point count censuses*

366 During the first week of July 2011-2013, we conducted a point-count census of birds
367 across clean and contaminated sites (Fig. 5). Each count lasted five minutes, with census points
368 located at approximately 100m intervals. At each census point, we classified the habitat as being
369 agricultural, grassland, deciduous forest or coniferous forest, and estimated ground coverage by
370 these different habitats (to the nearest 10%) within a distance of 50 m. In total, we collected 1100
371 5-min point counts (2011: N = 300; 2012: N = 400; 2013: N = 400). The census points were the
372 same in all years, except for 2011, when 100 fewer counts were conducted due to restrictions in
373 access. The relationship between radiation and abundance did not qualitatively change if we
374 restricted the analyses to the 300 points where the counts were conducted in all three years
375 (results not shown). At each census point, we recorded radiation levels using a hand-held
376 dosimeter at ground level (Model: Inspector, SE International, Inc., Summertown, TN, USA).
377 We also recorded the geographic coordinates and altitude (using a GPS), cloud cover at the start
378 of each point count (to the nearest eighth), temperature (degrees Celsius), and wind force
379 (Beaufort). For each census point we recorded time of day at the start of the count (to the nearest
380 minute) and included it in the analyses as an explanatory variable. As activity levels of birds
381 peak in the morning and, to a lesser extent, in the evening, we also included time squared in our
382 analyses. APM conducted all censuses, thus preventing any issue due to inter-observer
383 variability. All the nests that we inspected during 2012-2013 would be fledged by the time we
384 conducted our census. Thus, no difference among years would be expected based on differences
385 in the timing of reproduction.

386 In a second set of analyses, we analyzed the probability of observing a juvenile barn
387 swallow (as identified from the short tail streamers and the pale coloration, using binoculars) as a

388 function of environmental radiation levels, as well as the same predictors that we included in the
389 analysis of barn swallow abundance. We also included the local abundance of adult barn
390 swallows as a predictor in the analysis, as juvenile barn swallows are the offspring of the adult
391 barn swallows present.

392

393 *Statistical analysis*

394 For the analysis of genetic integrity of nestlings we used general linear mixed models
395 (GLMMs) where we included radiation exposure (either log-transformed radioactivity of the nest
396 material or radiation dose as inferred from the TLDs) as a covariate, and the nest of origin as a
397 random effect. In both analyses we included duration of exposure as a covariate. Degrees of
398 freedom were estimated using the Kenward-Roger approximation. All analyses were performed
399 in SAS 9.3 (SAS Inc., Cary, NC).

400 In the analysis of the abundance of barn swallows, we used generalized linear models,
401 assuming a Poisson distribution of count data. As predictor variables, we included \log_{10} -
402 transformed radiation and all potentially confounding variables listed above. In addition, we
403 included temperature, cloud cover, wind, time of day and time of day squared, the latter to
404 account for the fact that bird activity has a peak during early morning and a second, milder peak
405 in the afternoon. We also included radiation level squared to account for non-linear relationships
406 between species richness and abundance, respectively, and radiation. These analyses were all
407 implemented in the statistical software JMP (SAS Institute Inc., 2012). In the analyses of the
408 proportion of swallows being juveniles, we relied on general linear models with binomially
409 distributed data and a logit link function.

410

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416 swallow nests, often on the very top of their house door. This paper is dedicated to all those that
417 had to leave their beloved houses – and barn swallows.

418

419 **Authors Contribution**

420 A.B.A, A.P.M. and T.A.M. conceived the research; A.B.A., K.K., W.K., H.S., E.A.,
421 A.P.M. and T.A.M. collected data in the field; A.B.A. and S.O. performed the lab assays; A.B.A.
422 and D.J.T. collected and analyzed spectra; A.B.A. and A.P.M. performed statistical analyses;
423 A.B.A. wrote the first version of the paper; all authors commented on the manuscript.

424

425 **Competing financial interests**

426 The Authors declare no competing financial interests. Correspondence and requests for
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428 (mousseau@sc.edu).

429

430 **References**

- 431 1. Yasunari, T. J. *et al.* Cesium-137 deposition and contamination of Japanese soils due to
432 the Fukushima nuclear accident. *PNAS* **108**, 19530–19534 (2011).
- 433 2. Zheng, J. *et al.* Isotopic evidence of plutonium release into the environment from the
434 Fukushima DNPP accident. *Sci Rep* **2**, 304 (2012).
- 435 3. Stohl, A., Seibert, P. & Wotawa, G. The total release of xenon-133 from the Fukushima
436 Dai-ichi nuclear power plant accident. *J Environ Radioact* **112**, 155–159 (2012).
- 437 4. Chino, M. *et al.* Preliminary estimation of release amounts of ¹³¹I and ¹³⁷Cs accidentally
438 discharged from the Fukushima Daiichi nuclear power plant into the atmosphere. *J Nucl
439 Sci Technol* **48**, 1129–1134 (2011).
- 440 5. Terada, H., Katata, G., Chino, M. & Nagai, H. Atmospheric discharge and dispersion of
441 radionuclides during the Fukushima Dai-ichi Nuclear Power Plant accident. Part II:
442 verification of the source term and analysis of regional-scale atmospheric dispersion. *J
443 Environ Radioact* **112**, 141–154 (2012).
- 444 6. Stohl, A. *et al.* Xenon-133 and caesium-137 releases into the atmosphere from the
445 Fukushima Dai-ichi nuclear power plant: determination of the source term, atmospheric
446 dispersion, and deposition. *Atmos. Chem. Phys. Discuss.* **11**, 28319–28394 (2011).
- 447 7. Buesseler, K. O. *et al.* Fukushima-derived radionuclides in the ocean and biota off Japan.
448 *PNAS* **109**, 5984–5988 (2012).
- 449 8. Bailly du Bois, P. *et al.* Estimation of marine source-term following Fukushima Dai-ichi
450 accident. *J Environ Radioact* **114**, 2–9 (2012).
- 451 9. Steinhauser, G., Brandl, A. & Johnson, T. E. Comparison of the Chernobyl and
452 Fukushima nuclear accidents: A review of the environmental impacts. *Sci Total Environ*
453 **470-471**, 800–817 (2014).
- 454 10. Kobayashi, T., Nagai, H., Chino, M. & Kawamura, H. Source term estimation of
455 atmospheric release due to the Fukushima Dai-ichi Nuclear Power Plant accident by
456 atmospheric and oceanic dispersion simulations: Fukushima NPP Accident Related. *J
457 Nucl Sci Technol* **50**, 255–264 (2013).
- 458 11. Beresford, N. A. & Copplestone, D. Effects of ionizing radiation on wildlife: What
459 knowledge have we gained between the Chernobyl and Fukushima accidents? *Integr
460 Environ Assess Manag* **7**, 371–373 (2011).
- 461 12. Kinoshita, N. *et al.* Assessment of individual radionuclide distributions from the
462 Fukushima nuclear accident covering central-east Japan. *PNAS* **108**, 19526–19529 (2011).
- 463 13. Hayama, S.-I. *et al.* Concentration of Radiocesium in the Wild Japanese Monkey (*Macaca
464 fuscata*) over the First 15 Months after the Fukushima Daiichi Nuclear Disaster. *PLoS
465 ONE* **8**, e68530 (2013).
- 466 14. Yamashiro, H. *et al.* Effects of radioactive caesium on bull testes after the Fukushima
467 nuclear plant accident. *Sci Rep* **3**, 2850 (2013).
- 468 15. Buesseler, K. O. Fishing for Answers off Fukushima. *Science* **338**, 480–482 (2012).
- 469 16. Ishida, K. (Contamination of Wild Animals: Effects on Wildlife in High Radioactivity
470 Areas of the Agricultural and Forest Landscape) *Agricultural Implications of the
471 Fukushima Nuclear Accident* [Nakanishi, T. M. & Tanoi, K. (eds)] [119–129] (Springer,
472 New York, 2013).
- 473 17. Higaki, T., Higaki, S., Hirota, M., Akita, K. & Hasezawa, S. Radionuclide Analysis on
474 Bamboos following the Fukushima Nuclear Accident. *PLoS ONE* **7**, e34766 (2012).
- 475 18. Yoshihara, T., Matsumura, H., Hashida, S.-N. & Nagaoka, T. Radiocesium

- 476 contaminations of 20 wood species and the corresponding gamma-ray dose rates around
 477 the canopies at 5 months after the Fukushima nuclear power plant accident. *J Environ*
 478 *Radioact* **115C**, 60–68 (2012).
- 479 19. Kuroda, K., Kagawa, A. & Tonosaki, M. Radiocesium concentrations in the bark,
 480 sapwood and heartwood of three tree species collected at Fukushima forests half a year
 481 after the Fukushima Dai-ichi nuclear accident. *J Environ Radioact* **122**, 37–42 (2013).
- 482 20. Hiyama, A. *et al.* The biological impacts of the Fukushima nuclear accident on the pale
 483 grass blue butterfly. *Sci Rep* **2**, 570 (2012).
- 484 21. Hiyama, A. *et al.* The Fukushima nuclear accident and the pale grass blue butterfly:
 485 evaluating biological effects of long-term low-dose exposures. *BMC Evol Biol* **13**, 168
 486 (2013).
- 487 22. Fujita, Y., Yoshihara, Y., Sato, I. & Sato, S. Environmental radioactivity damages the
 488 DNA of earthworms of Fukushima Prefecture, Japan. *Eur J Wildlife Res* **60**, 145–148
 489 (2014).
- 490 23. Ochiai, K. *et al.* Low blood cell counts in wild Japanese monkeys after the Fukushima
 491 Daiichi nuclear disaster. *Sci Rep* **4**, 5793 (2014).
- 492 24. Uematsu, S., Uematsu, K., Lavers, J. L. & Congdon, B. C. Reduced vitamin A (retinol)
 493 levels indicate radionuclide exposure in Streaked Shearwaters (*Calonectris leucomelas*)
 494 following the 2011 Fukushima nuclear accident. *Ecol Indic* **43**, 244–251 (2014).
- 495 25. Mousseau, T. A. & Møller, A. P. Genetic and ecological studies of animals in Chernobyl
 496 and Fukushima. *J Hered* **105**, 704–709 (2014).
- 497 26. Bonisoli Alquati, A. *et al.* DNA damage in barn swallows (*Hirundo rustica*) from the
 498 Chernobyl region detected by use of the comet assay. *Comp Biochem Phys C* **151**, 271–
 499 277 (2010).
- 500 27. Møller, A. P., Mousseau, T. A., De Lope, F. & Saino, N. Elevated frequency of
 501 abnormalities in barn swallows from Chernobyl. *Biol Lett* **3**, 414–417 (2007).
- 502 28. Møller, A. P., Bonisoli-Alquati, A. & Mousseau, T. A. High frequency of albinism and
 503 tumours in free-living birds around Chernobyl. *Mutat Res-Gen Tox En* **757**, 52–59 (2013).
- 504 29. Møller, A. P., Bonisoli-Alquati, A., Rudolfson, G. & Mousseau, T. A. Chernobyl birds
 505 have smaller brains. *PLoS ONE* **6**, e16862 (2011).
- 506 30. Møller, A. P., Erritzøe, J., Karadas, F. & Mousseau, T. A. Historical mutation rates predict
 507 susceptibility to radiation in Chernobyl birds. *J Evol Biol* **23**, 2132–2142 (2010).
- 508 31. Møller, A. P. & Mousseau, T. A. Determinants of interspecific variation in population
 509 declines of birds after exposure to radiation at Chernobyl. *J Appl Ecol* **44**, 909–919 (2007).
- 510 32. Møller, A. P., Bonisoli Alquati, A., Rudolfson, G. & Mousseau, T. A. Elevated Mortality
 511 among Birds in Chernobyl as Judged from Skewed Age and Sex Ratios. *PLoS ONE* **7**,
 512 e35223 (2012).
- 513 33. Møller, A. P. *et al.* Abundance of birds in Fukushima as judged from Chernobyl. *Environ*
 514 *Pollut* **164**, 36–39 (2012).
- 515 34. Møller, A. P. *et al.* Differences in effects of radiation on abundance of animals in
 516 Fukushima and Chernobyl. *Ecol Indic* **24**, 75–81 (2013).
- 517 35. Møller, A. P. *et al.* Condition, reproduction and survival of barn swallows from Chernobyl.
 518 *J Anim Ecol* **74**, 1102–1111 (2005).
- 519 36. Anspaugh, L. R. Doses to members of the general public and observed effects on biota:
 520 Chernobyl Forum update. *J Environ Radioact* **96**, 13–19 (2007).
- 521 37. Galván, I. *et al.* Chronic exposure to low-dose radiation at Chernobyl favours adaptation

- 522 to oxidative stress in birds. *Funct Ecol* **28**, 1387–1403 (2014).
- 523 38. Alonso-Álvarez, C., Bertrand, S., Faivre, B. & Sorci, G. Increased susceptibility to
524 oxidative damage as a cost of accelerated somatic growth in zebra finches. *Funct Ecol* **21**,
525 873–879 (2007).
- 526 39. Moreno-Rueda, G., Redondo, T., Trenzado, C. E., Sanz, A. & Zúñiga, J. M. Oxidative
527 Stress Mediates Physiological Costs of Begging in Magpie (*Pica pica*) Nestlings. *PLoS*
528 *ONE* **7**, e40367 (2012).
- 529 40. Møller, A. P., Mousseau, T. A., Lynn, C., Ostermiller, S. & Rudolfson, G. Impaired
530 swimming behaviour and morphology of sperm from barn swallows *Hirundo rustica* in
531 Chernobyl. *Mutat Res-Gen Tox En* **650**, 210–216 (2008).
- 532 41. Møller, A. P. & Mousseau, T. A. Mutation and sexual selection: A test using barn
533 swallows from Chernobyl. *Evolution* **57**, 2139–2146 (2003).
- 534 42. Møller, A. P., Karadas, F. & Mousseau, T. A. Antioxidants in eggs of great tits *Parus*
535 *major* from Chernobyl and hatching success. *J Comp Physiol B* **178**, 735–743 (2008).
- 536 43. Taira, Y. *et al.* Environmental contamination and external radiation dose rates from
537 radionuclides released from the Fukushima nuclear power plant. *Radiat Prot Dosim* **151**,
538 537–545 (2012).
- 539 44. Garnier-Laplace, J. M., Beaugelin-Seiller, K. & Hinton, T. G. Fukushima wildlife dose
540 reconstruction signals ecological consequences. *Environ Sci Technol* **45**, 5077–5078
541 (2011).
- 542 45. Strand, P. *et al.* Assessment of Fukushima-Derived Radiation Doses and Effects on
543 Wildlife in Japan. *Environ Sci Technol Lett* **1**, 198–203 (2014).
- 544 46. Ulanovsky, A., Pröhl, G. & Gómez-Ros, J. M. Methods for calculating dose conversion
545 coefficients for terrestrial and aquatic biota. *J Environ Radioact* **99**, 1440–1448 (2008).
- 546 47. Møller, A. P. & Mousseau, T. A. Efficiency of bio-indicators for low-level radiation under
547 field conditions. *Ecol Indic* **11**, 424–430 (2011).
- 548 48. Smith, J. T. Is Chernobyl radiation really causing negative individual and population-level
549 effects on barn swallows? *Biol Lett* **4**, 63–64 (2008).
- 550 49. ICRP. Environmental Protection: the Concept and Use of Reference Animals and Plants.
551 ICRP Publication 108. *Ann ICRP* **38**, (2008).
- 552 50. Howard, B. J. *et al.* The IAEA handbook on radionuclide transfer to wildlife. *J Environ*
553 *Radioact* **121**, 55–74 (2013).
- 554 51. Garnier-Laplace, J. M. *et al.* Are radiosensitivity data derived from natural field
555 conditions consistent with data from controlled exposures? A case study of Chernobyl
556 wildlife chronically exposed to low dose rates. *J Environ Radioact* **121**, 12–21 (2013).
- 557 52. Møller, A. P. & Mousseau, T. A. The effects of natural variation in background
558 radioactivity on humans, animals and other organisms. *Biol Rev* **88**, 226–254 (2012).
- 559 53. Shen, W., Tang, K., Zhu, H. & Liu, B. New Advances in LiF: Mg, Cu, P TLD (GR-200A).
560 *Radiat Prot Dosim* **100**, 357–360 (2002).
- 561 54. Bacci, C., D'Angelo, L., Furetta, C. & Giancola, S. Comprehensive study on LiF: Cu, Mg,
562 P (GR-200 A). *Radiat Prot Dosim* **47**, 215–218 (1993).
- 563 55. Singh, N. P. *et al.* Abundant alkali-sensitive sites in DNA of human and mouse sperm.
564 *Exp Cell Res* **184**, 461–470 (1989).
- 565 56. Rosenberger, A. *et al.* Validation of a fully automated COMET assay: 1.75 million single
566 cells measured over a 5 year period. *DNA Repair* **10**, 322–337 (2011).
- 567 57. Tice, R. R. *et al.* Single cell gel/comet assay: guidelines for in vitro and in vivo genetic

- 568 toxicology testing. *Environ Mol Mutag* **35**, 206–221 (2000).
- 569 58. Cotellet, S. & Ferard, J. Comet assay in genetic ecotoxicology: A review. *Environ Mol*
- 570 *Mutag* **34**, 246–255 (1999).
- 571
- 572

573 **Figure captions**

574

575 Figure 1. Radiation measurements and genetic damage. The relationship between genetic
576 damage of nestlings and (a) the activity concentrations of the nest material (Bq/kg d.w.,
577 summing activities of ^{134}Cs and ^{137}Cs) or (b) external radiation dose rate, as measured by the TLD
578 ($\mu\text{Gy h}^{-1}$). The lines are simple regression lines interpolated to the log-transformed data.

579

580 Figure 2. Barn swallows abundance and radioactive contamination. The abundance of barn
581 swallows declined with increasing levels of radioactive contamination as measured during our
582 multi-year point-count census.

583

584 Figure 3. Age ratio of barn swallows and radioactive contamination. The proportion of barn
585 swallows being juveniles declined with increasing levels of radioactive contamination.

586

587 Figure 4. Locations of sampling sites. Locations of the sixteen nests used in the analyses of the
588 relationship between contamination levels and DNA damage of the nestlings. Each location may
589 correspond to more than one nest sampled. Contamination levels are derived from official data
590 from the Japanese Ministry of Education, Culture, Sports, Science and Technology (MEXT), and
591 used to interpolate a map of contamination at 1-m height. The map was created using ArgGis
592 v10.2 (Environmental Systems Research Institute, Redlands, CA).

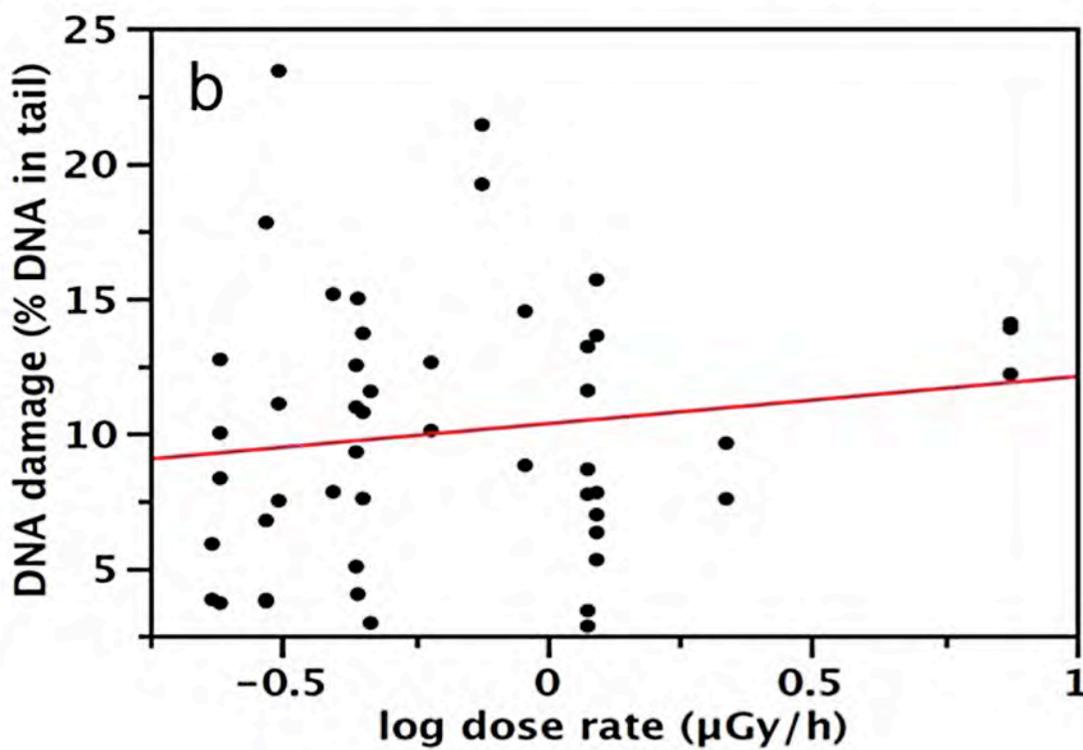
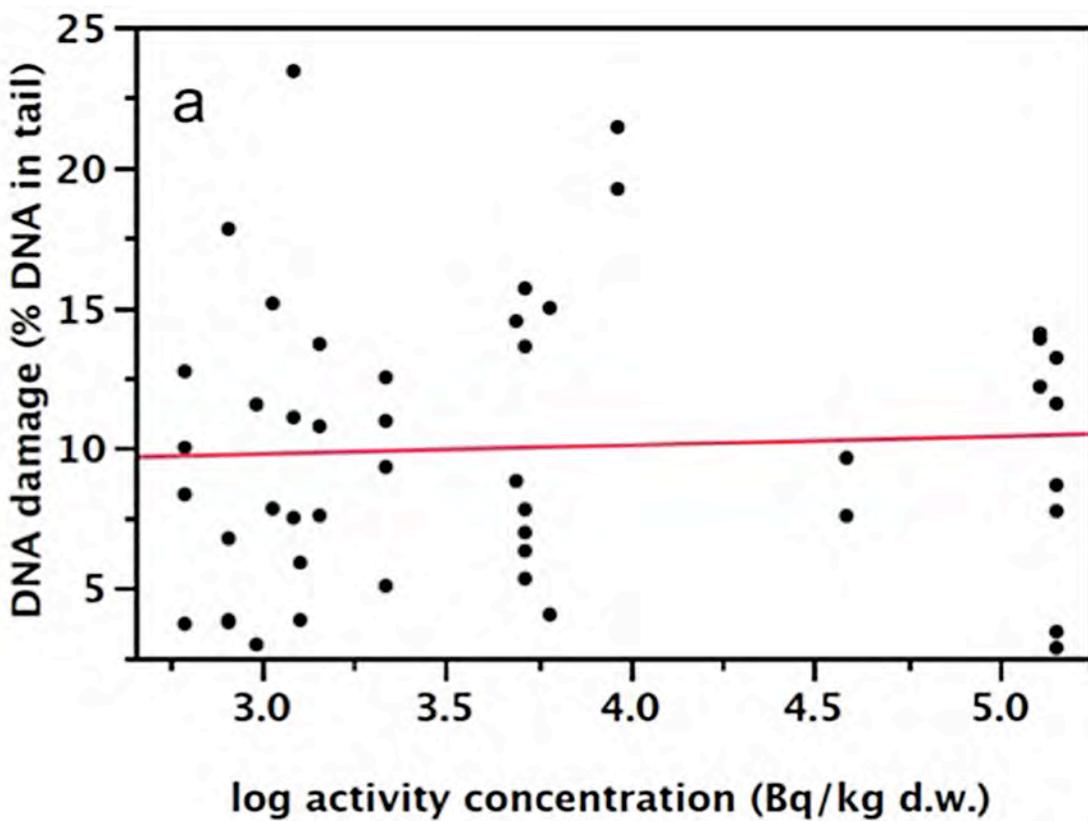
593

594 Figure 5. Locations of census sites during 2011-2013. Contamination levels are derived from
595 official data from the Japanese Ministry of Education, Culture, Sports, Science and Technology
596 (MEXT), and used to interpolate a map of contamination at 1-m height. The map was created
597 using ArgGis v10.2 (Environmental Systems Research Institute, Redlands, CA).

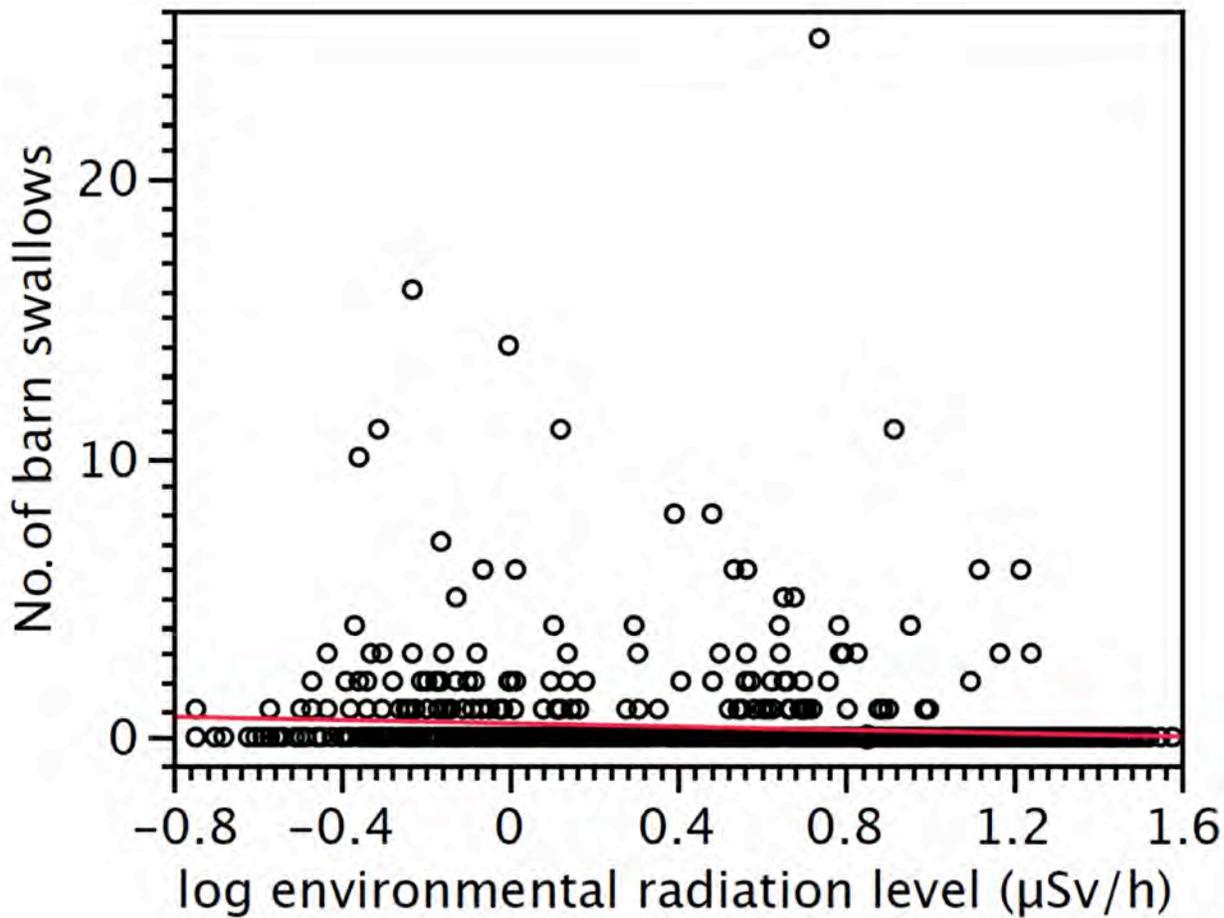
598

599

600 Figure 1



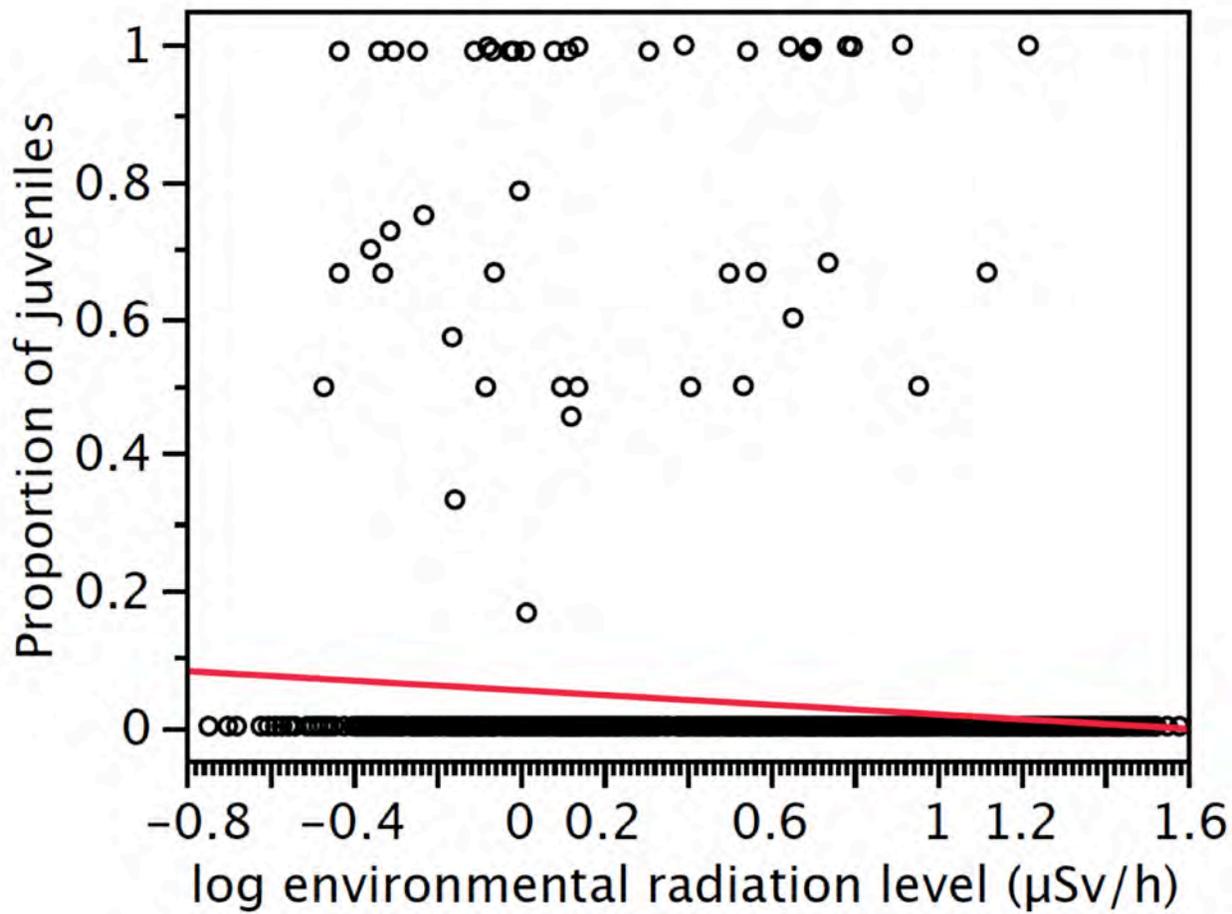
602 Figure 2



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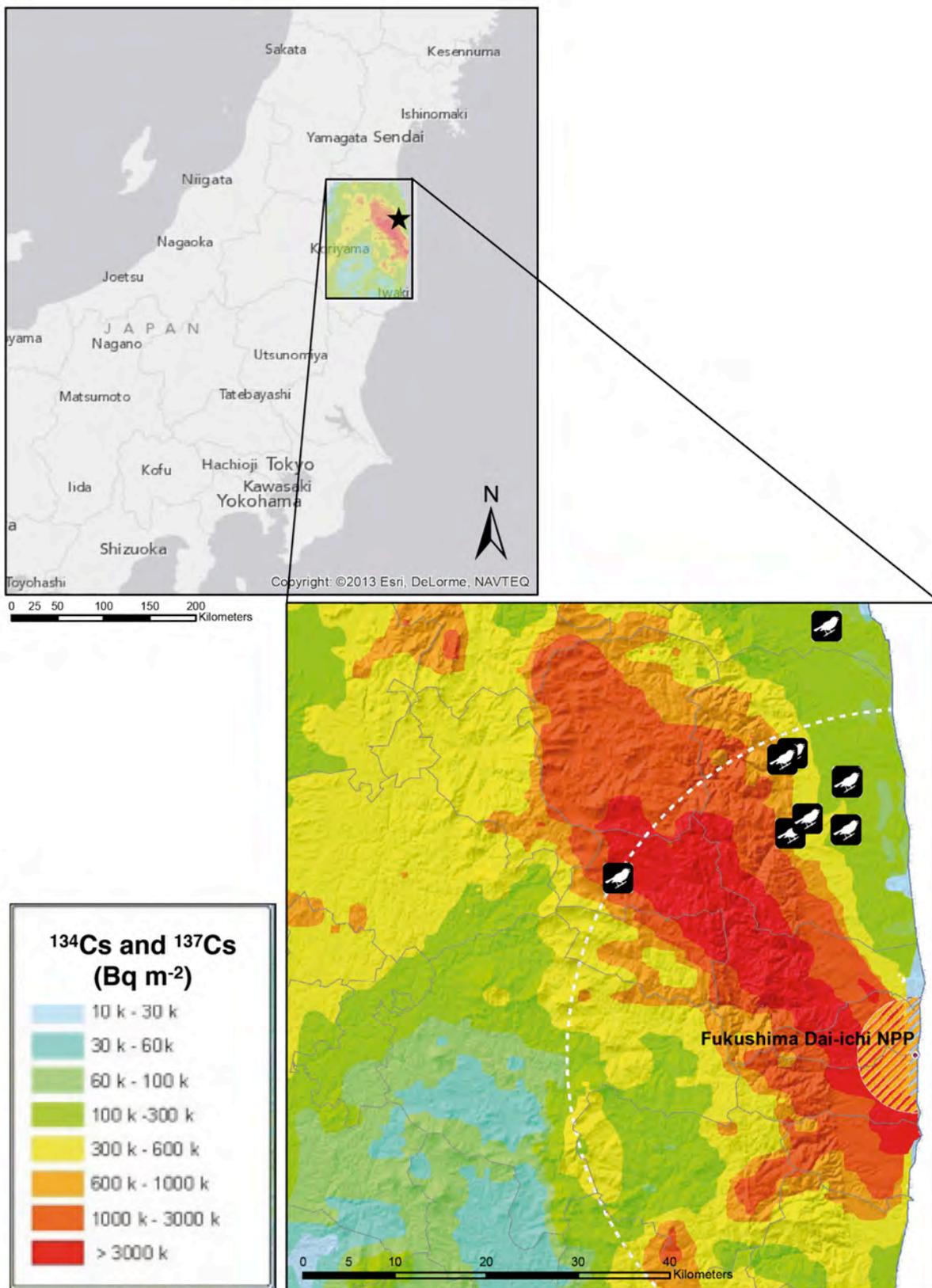
605 **Figure 3**



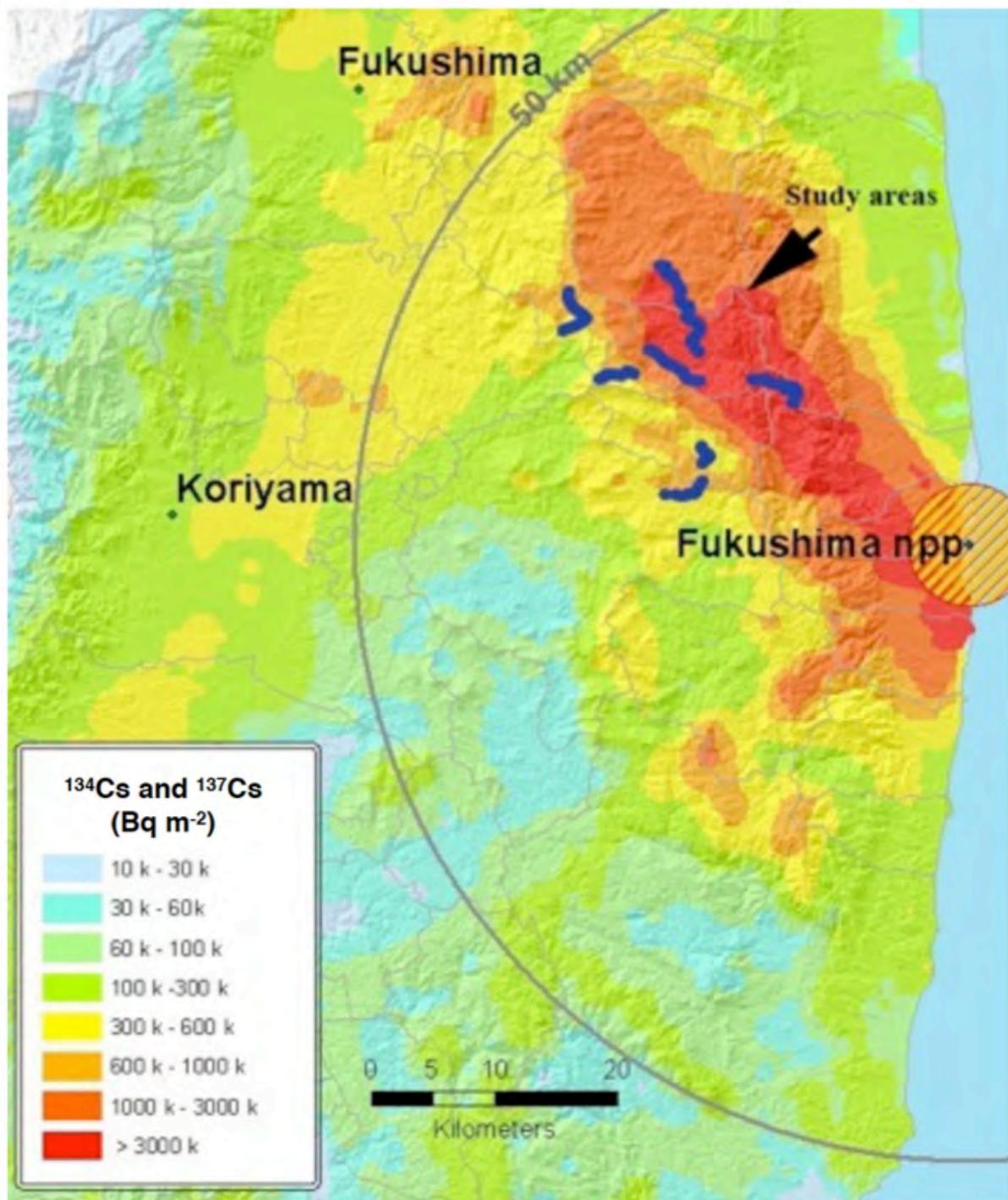
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608 **Figure 4**



610 **Figure 5**



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612